Profil Proporsi Streptococcus mutans pada Beberapa Lesi Karies dengan Tingkat Keparahan Berbeda

(Streptococcus mutans Proportion Profile in Several Caries Lesions with Different Severity Levels)

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Abstrak

Karies gigi adalah penyakit menular bakteri umum yang memengaruhi struktur gigi, dengan Streptococcus mutans dikaitkan dengan tingkat keparahannya. Penelitian ini bertujuan untuk menilai distribusi proporsi S. mutans di berbagai tingkat lesi karies sebagai faktor prognostik potensial. Desain kelompok kontrol pasca-tes saja dilakukan pada 33 pasien yang terdaftar di RSGM UMY yang memenuhi kriteria sampel dengan gigi bebas karies, gigi berlubang berat, pulpitis reversibel, dan nekrosis pulpa. Sampel DNA diekstraksi dan dianalisis menggunakan teknik polymerase chain reaction (PCR) dan analisis densitometri semi kuantitatif menggunakan Image-J. Analisis statistik dilakukan untuk membandingkan proporsi S. mutans di berbagai kategori lesi. Studi ini menemukan variasi yang tidak signifikan dalam proporsi S. mutans (p>0.05) di antara kelompok yang diamati, dengan tingkat populasi tertinggi diamati pada kelompok ICDAS V (34,53±23,09), diikuti oleh kelompok nekrosis pulpa (24,41±15,48), kelompok pulpitis reversibel (11,14±1,36), dan kelompok gigi bebas karies (6,95±2,57). Sebaliknya, proporsi populasi terendah ditemukan dalam kondisi pulpitis bebas karies dan reversibel. Meskipun S. mutans dikaitkan dengan karies, keberadaannya saja tidak sepenuhnya menjelaskan dan berkorelasi dengan tingkat keparahan lesi.

Kata kunci: Karies, Karies bebas, Proporsi bakteri, Streptococcus mutans

Abstract

Dental caries is a common bacterial infectious disease that affects tooth structure, with Streptococcus mutans being associated with its severity. This study aimed to assess the distribution of S. mutans proportions across various levels of carious lesions as a potential prognostic factor. A post-test-only control group design was conducted on 33 patients registered at RSGM UMY who fulfilled the sample criteria with caries-free teeth, severe cavities, reversible pulpitis, and pulp necrosis. DNA samples were extracted and analyzed using the polymerase chain reaction (PCR) technique and semi-quantitative densitometry analysis using Image-J. Statistical analysis was performed to compare S. mutans proportions across different lesion categories. The study found non-significant variations in S. mutans proportions (p>0.05) among the observed groups, with the highest population levels observed in the ICDAS V group (34.53±23.09), followed by the pulp necrosis group (24.41±15.48), reversible pulpitis group (11.14±1.36), and the caries-free teeth group (6.95±2.57). In contrast, the lowest proportions of the population were found in caries-free and reversible pulpitis conditions. Although S. mutans is associated with caries, its presence alone does not fully explain and correlate to the lesion severity.

Keywords: Bacterial proportion, Caries, free caries, Streptococcus mutans

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Dental caries is a widespread global oral health issue that affects a significant coverage within the population, with prevalence rates of 80-90% in various communities¹. In Indonesia, the prevalence of dental caries is notably high, reported at 88.8%, according to the 2018 Basic Health Results published by the Ministry of Health². Dental caries is a chronic bacterial infectious disease that involves the tooth structure, affects both hard and soft tissues, and can progress gradually to destroy the integrity of the tooth³. The development of caries involves complex interactions among host factors, dietary habits, time, and microorganisms. Tooth decay is a dental condition where the hard tissues of the teeth break down due to bacteria, substrates, and host factors⁴. The term 'caries,' which means decay, is affected by various factors like the presence of decay-causing bacteria such as Streptococcus mutans, the intake of sugary foods, and how long they are in contact with the teeth⁵. Cavities may develop when a sticky film of bacteria, called plaque, builds up on the teeth and interacts with sugars to create acid, which damages the teeth by removing minerals. If the loss of minerals continues without enough repair, holes in the

teeth, known as cavities, can develop and cause permanent harm to the tooth structure⁶.

The International Caries Detection and Assessment System (ICDAS) is a globally recognized system for assessing dental caries and tracking their frequency and severity. ICDAS provides a framework for improved clinical management of caries by classifying the condition from the healthy teeth (ICDAS code 0) to severe decay (ICDAS code 6)7. The system identifies various stages of caries development, including initial visual changes in enamel (D1), clear visual changes (D2), early enamel damage (D3), localized enamel damage with dentin shadow (D4), and extensive decay with exposed dentin (D5)⁸. The most severe stage, ICDAS code 6, indicates deep cavities that involve more than half of the tooth surface and may potentially reach the pulp⁹.

Pulp disease encompasses various conditions that affect the dental pulp; normal pulp is classified as clinically sensitive yet asymptomatic. The most prevalent cause of pulp disease is bacterial invasion through the dentin, typically resulting from untreated caries (tooth decay). Inflammation of the pulp, known as pulpitis, can be categorized as reversible or irreversible. Reversible pulpitis is caused by mild inflammation that resolves once the irritant is removed. Irreversible pulpitis involves persistent inflammation that cannot return to a normal state and is often accompanied by severe pain that may radiate to adjacent teeth¹⁰. Understanding the factors behind tooth decay is vital for creating effective strategies to prevent and treat dental problem issues.

Streptococcus mutans is а microorganism component among the most common bacterium associated with tooth decay. S mutants also belong to acidogenic and acidic pathogens harboring the capacity to produce organic acids through sugar metabolism, thereby contributing to the cariogenic process¹¹. Streptococcus mutans is a gram-positive bacterium that can survive with or without oxygen and is known as a primary pathogen associated with dental caries¹². It belongs to the genus Streptococcus and can be classified into different variations, with type C being the most common in humans¹³. The bacterium plays a crucial role in causing cavities by breaking down sugars into lactic acid, lowering the pH, and destructing the enamel's hard tissue. S mutants can also produce sugars outside their cells from sucrose, promoting biofilm formation and helping it stick to tooth surfaces¹⁴.

The harmful effects of *S* mutants are due to specific factors like Gtf and Gbp, which help in producing sticky substances. These factors contribute to the synthesis of sticky glucans, which form a robust extracellular matrix in dental biofilms¹⁵. Continuous sucrose intake by the host promotes an acidic environment that favors the growth of *S* mutants while inhibiting competing bacteria. This change leads to the production of organic acids, which lowers the pH and starts the formation of cavities. Understanding *S* mutant's role in dental caries is crucial for developing effective prevention and treatment strategies.

Although S. mutans is frequently regarded as the primary pathogen, its prevalence in carious lesions compared to caries-free conditions varies, with several studies reporting inconsistent prevalence rates^{16,17}. Previous study by Frakgou et al. reported variations in the prevalence of S. mutans between caries-free and caries-active children. In this study, which included 51 caries-active and 46 caries-free children aged 3 to 13 years, S. mutans was detected in 66% of samples¹⁷. Similarly, a study by Acevedo et al. examined the prevalence of S. mutans in children with and without caries, reporting its presence in 10 out of 30 children with caries (33%) and 6 out of 18 caries-free children (33%) aged 2 to 19 years, with no significant difference between the groups¹⁸. Another finding from Lima et al., and Thimmegowda reported a low prevalence rate of S. mutans in non-cavity-free teeth^{19,20}. All of these reports suggest that while S. mutans is commonly associated with dental caries, its prevalence varies across different populations and does not always correlate directly with caries status.

Differences in the proportion of *S. mutans* in carious lesions are believed to influence the severity of dental caries. Some reports indicate that since *S. mutans* possesses cariogenic properties, caries still can develop in its absence; on the other hand, *S. mutans* can also be present without any tooth decay development²¹. Therefore, this study aims to evaluate the pathogenic proportion of *Streptococcus mutans* at several levels of carious lesion severity as a pathogenic prognostic factor. The study will involve sampling from different carious lesion conditions, including active carious lesion (ICDAS V), necrotizing caries, reversible pulpitis, and caries-free states, to observe the proportion of *S. mutans* for dental caries development.

METHODS

This research is a study that observes subjects in a controlled environment using a posttest-only control group design. The study included patients with specific dental conditions, such as caries-free teeth, teeth with severe cavities, reversible pulpitis, and pulp necrosis, registered at RSGM UMY. The research protocol was approved by the Ethical Committees of the Faculty of Medicine and Health Sciences Universitas Muhammadiyah Yogyakarta 090/EC-KEPK FKIK UMY/II/2024. Samples were collected intentionally to ensure a balanced representation, resulting in 33 samples evenly divided based on lesion criteria. The sampling process involved using a swab to collect *Streptococcus mutans* bacteria from the surfaces of both caries-free teeth and teeth with cavities.

The inclusion criteria for this study comprised subjects with caries-free teeth, caries classified as code 5 according to the International Caries Detection and Assessment System (ICDAS), caries with reversible pulpitis, and caries with pulp necrosis. Additionally, only patients who provided informed consent and those attending their first visit for caries treatment were included. The exclusion criteria encompassed patients with a poor Oral Hygiene Index-Simplified (OHI-S) score (3.1–6), individuals with disabilities, those attending follow-up visits after their initial consultation such as cases where caries removal had already been performed and patients who had completed their first-visit treatment.

The analysis process was conducted at a laboratory. It involved amplifying DNA using a Polymerase Chain Reaction (PCR) technique to detect the presence of *S. mutans*, followed by gel analysis and documentation to visualize the results.

Bacterial DNA were extracted using TRNzol Universal Reagent (TIANGEN BIOTECH) according to the manufacturer's protocols and used as templates for downstream analysis. Bacterial samples were homogenized in TRNzol Universal Reagent, followed by the addition of chloroform and centrifugation to separate the aqueous phase. The DNA was precipitated using isopropanol, washed with 75% ethanol, and resuspended in nuclease-free water. The quality and concentration of the extracted DNA were assessed using a NanoVue Plus (Biochrom Ltd., Cambridge, UK) before further analysis.

For gene detection analysis, 5 μ L DNA template was mixed with 25 μ L 2x MyTaq HS Red Mix (Bioline, UK) and 18 μ L nuclease free water (NFW) along with the indicated specific primers (Table 1)²², following the conditions listed in Table 1.

Table 1. List primer and PCR setting for S. mutans					
Primer	Temperature (°C)	Time	Cycle		
Forward 5'-ACTACACTITCGGGTGGCTTGG -3'	95	4 min	1		
Reverse 5'-CAGTATAAGCGCCAGTTTCATC -3'	95	1 min			
	55	1 min	35		
	72	1 min			
	72	10 min	1		
	٨	Hold	1		

Amplification products were visualized by electrophoresis and densitometry analysis according to Adiningrat et al., with slight modifications in volume²³. 8 µL PCR products from the gene detection analysis were mixed with 2 µL Novel Juice (dye-containing loading buffer, GeneDireX, USA) and loaded into a 1% agarose gel with 1 µL GelRed (Biotium, Canada). Electrophoresis gel analysis was then carried out at 50 V for 50–60 min. The gel was visualized with a UV (Genedirex, Transilluminator USA) after electrophoresis.

Semi-quantitative densitometry analysis using ImageJ was performed following the method described by Adiningrat et al. The analysis began by importing the gel electrophoresis results and converting the image to grayscale format. The Rectangle Tool was then used to select the band area for analysis, ensuring that all bands within the same gel were measured using a consistent Region of Interest (ROI) to maintain accuracy. The Measure feature (Analyze → Measure) was utilized to obtain the area under the curve value, which

represents band intensity. Band intensity was then normalized against the control band. Finally, data interpretation was conducted by comparing intensity values between samples to assess the relative amount of the target, with further statistical analysis performed using Excel²³.

The normality of the data will be assessed using the Shapiro-Wilk test. If the data are normally distributed, an independent t-test will be performed to identify differences in S. mutans proportions at different levels of caries severity. Statistical analysis will be conducted using IBM SPSS Statistics version 25 (IBM Corp., Armonk, NY, USA) to ensure the validity and reliability of the results.

RESULTS

All samples used in this study underwent semi-quantitative PCR testing with densitometry to determine the presence of a DNA fragment and to measure the intensity of the DNA band. The results of this test are expressed as optical density (OD), as shown in Figure 1 and Table 2.



Figure 1. Gel analysis visualization of PCR products from 12 samples with S. mutans primer (517 bp) and 16s primer (69 bp)

Table 2	2 . S	emi-qu	antitative	PCR	test	result
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Sample	Depetition	Dereentage	Magn	St Dov	٥٢
sample	Repention	Ferceniuge%	mean	31.Dev	3E
	A1	2.37			
FREE CARIES	A3	11.25	6.95	4.446617	2.567255
	A5	7.23			
	B1	80.55			
ICDAS V	B4	14.81	34.53	39.98677	23.08637
	B6	8.24			
	C1	11.64			
P. REVER	C6	13.20	11.14	2.350234	1.356908
	C3	8.58			
	D3	9.96			
NEKROSIS	D4	7.92	24.41	26.81423	15.4812
	D5	55.35			





The data presented in Figure 1 results from a densitometry analysis conducted on 12 samples. All samples showed the presence of bacteria when analyzed with the 16S primer (69 bp). However, specific signals were detected in only three samples when assessed with the *S. mutant* 'sspecific primer (517 bp): two from the ICDAS V category and one from the pulp necrosis category. The table summarizes the results of the densitometry calculations (Table 2)

The data presented in Table 2 calculates the percentage of densitometry results, indicating that the highest rate of *S. mutans* is found in sample B1 (ICDAS V) at 80.55%, while the lowest percentage is observed in the categories of cariesfree.

The boxplot, a graphical representation of the distribution of *S. mutants* across the four caries lesion categories, is shown in Figure 2. The range of data values is narrow for free caries and reversible pulpitis, with the median almost reaching 10%. The median for the ICDAS V group is around 45%, compared to about 30% for the pulp necrosis group.

To assess if the data follows a normal distribution, a normality test using the Shapiro-Wilk test will be performed on the dataset from Table 2 due to the sample size being less than 50.

The normality test results show a significance value (p > 0.05), indicating that the data follows a normal distribution. Using these results, a parametric test, such as the independent t-test, will be used to determine if there are notable variations in the population distribution across each sample category.

From Figure 2, it can be seen from the results of the independent sample t-test that the results of each test between categories are not significantly different.

DISCUSSION

The results of this study indicate a trend toward normal distribution in the statistical analysis of Streptococcus mutans proportions across different caries severity groups. However, no significant differences were observed between the control (caries-free) and target groups, including ICDAS V, reversible pulpitis, and pulp necrosis. These findings suggested that although S. mutans is commonly associated with the development of dental caries, the variable of its presence only may not be sufficient to differentiate the various carious lesion conditions. This aligns with the growing body of evidence indicating that caries development is multifactorial process involving complex а microbial interactions in a longitudinal process rather than the sole presence of S. mutans at a single point in time 24-26.

Dental caries is a complex pathological cross-interactions condition where amona bacteria, substrate, host, and time contribute to the process of enamel demineralization^{26,25} due to the increase of biofilm acidity. Among the various microbial factors involved in caries progression, S. mutans plays a crucial role due to its ability to thrive in acidic environments. Both acidogenicity and aciduric properties are important biochemical characteristics for the cariogenic capacity of S. mutans²⁷. On the other hand, the occurrence of dynamic microbial interaction may also exhibit microbial homeostasis that may control microbiome pathogenicity. Veillonella is able to use the lactic acid produced by S. mutans as a carbon source, thus acting as an "acid sink." This may prevent the biofilm from reaching a very low pH, which could reduce the ability of S. mutans to survive and contribute to caries progression¹⁶. As probiotics, some Lactobacillus species can also reduce the cariogenic activity of S. mutans. They compete for space and nutrients in the complex biofilm and produce antimicrobial metabolites that can inhibit the growth of *S. mutans,* thus reflecting on the internal pathogenicity control mechanism²⁹.

Although Streptococcus mutans is often linked to caries development, our study found that it did not consistently dominate in all cases, suggesting the involvement of other bacterial species during the caries development process since it is also inevitable existence. This result may relate to the previous report, which demonstrates the possible dynamic carious lesion development process by not only a single bacteria domination approach but also a collective bacterial approach within the carious lesion microbiome. For example, previous research conducted by Klein et al. revealed that other bacteria, such as Lactobacillus spp. and Actinomyces spp., may contribute to caries formation, especially under conditions where S. mutans is absent³⁰. Other studies have shown that bacteria other than S. mutans play a dominant role in caries Species Veillonella, development. from Lactobacillus, Bifidobacterium, Propionibacterium, Atopobium, and Actinomyces, along with non-S. mutans acidogenic streptococci, often at higher levels than S. mutans³¹. Furthermore, Marsh emphasized the importance of understanding the broader microbial ecosystem within the oral cavity, where interactions among different microbial species can influence caries development, even in the absence of S. mutans³². This indicates that dental caries could be dependent not only on a single type of bacteria but also on the complex interactions among various microbes in the dental biofilm.

The non-dominance of *S. mutans* in pulp necrosis and reversible pulpitis suggested a shift in the microbial landscape, with other bacterial groups such as Actinomyces spp., *Staphylococcus aureus, Firmicutes*, and Actinobacteria, becoming more prevalent ^{33–37}. These findings suggested a significant shift in the oral microbiome, where diverse acidogenic bacteria may play a more prominent role in the transition from early carious lesions to pulpitis. The observed variability in *S. mutans* presence among individuals underscores the complex interactions within the oral microbial community³⁸. This variability also presents a challenge in ensuring sample homogeneity which may influence data consistency.

Sample homogeneity remains a key challenge and limitation of this study. Variations in clinical conditions during the pre-analytical phase, including differences in cavity or caries status at initial sample collection, may have led to sample loss and compromised data consistency. To address this, further optimization of pre-analytical procedures is necessary to improve sample standardization and ensure data reliability. Additionally, expanding sample coverage is essential to enhance the study's representativeness and strengthen its external validity.

Although Streptococcus mutans is often considered as the main pathogen factor associated with dental caries, the findings from this study, corresponding with previous studies, suggested that its presence as normal oral flora did not solely and directly correlate with caries development at a single point of time³⁹. The absence of significant differences in the proportion of S. mutans among different dental carious lesion conditions also indicated the possibility that other microbial species and environmental factors could play an important role in dynamic caries development. These underline the complexity of the oral microbiome and highlight the need for a deeper understanding of the interactions between different bacterial populations toward carious lesion development. Further investigation is essential to elucidate the multi-factorial nature of carious and to refine prevention and better treatment strategies.

From a clinical perspective, if the findings of this study support the proposed hypothesis, they could contribute to minimizing the severity of dental caries. Early detection of *Streptococcus mutans* could be incorporated as part of a screening protocol, allowing for a preventive approach to reduce potential for increased severity of caries in the future.

ACKNOWLEDGMENTS

We would like to express our gratitude to the Universitas Muhammadiyah Yogyakarta for their financial support and to the DH-RIC MMT Laboratory UMY for their invaluable assistance in providing research tools, facilities, materials, and experimental support.

REFERENCES

- 1 Teshome A, Muche A, Girma B. Prevalence of Dental Caries and Associated Factors in East Africa, 2000– 2020: Systematic Review and Meta-Analysis. Front Public Health. 2021;9:645091.
- 2 Utami S, Pinastika R, Astuti NR, Adiningrat A. The Overview of Eating Patterns and Dental Caries Status of The Community of Pendul, Argorejo, Bantul Yogyakarta. Insisiva Dent J Maj Kedokt Gigi Insisiva. 2023;12(1):1–6.
- 3 Alsuraim BS, Han D-H. Effect of globalization on global dental caries trend. *Medicine (Baltimore)*. 2020;99(35):e21767.
- 4 Villhauer A, Lynch D, Postler T, Dawson D, Drake D. Mutans Streptococci and Lactobacilli: Colonization Patterns and Genotypic Characterization of Cariogenic Bacterial Species in American Indian Children. Front Dent Med. 2021;2:740900.
- 5 Mathur VP, Dhillon JK. Dental Caries: A Disease Which Needs Attention. Indian J Pediatr. 2018;85(3):202–206.
- 6 Adiningrat A, Kusmaharani H, Utami S, Ratna Astuti N. Evaluation of International Caries Detection and Assessment System

(ICDAS)-related caries severity among caries risk groups in Pendul district: An observational study. J Int Soc Prev Community Dent. 2020;10(4):498.

- 7 Schoilew K, Ueffing H, Dalpke A et al. Bacterial biofilm composition in healthy subjects with and without caries experience. J Oral Microbiol. 2019;11(1):1633194.
- 8 Dikmen B. ICDAS II CRITERIA (INTERNATIONAL CARIES DETECTION AND ASSESSMENT SYSTEM). J Istanb Univ Fac Dent. 2015;49(3):63.
- 9 Conrads G, About I. Pathophysiology of Dental Caries. In: Schwendicke F, Frencken J, Innes N (eds). Monographs in Oral Science. S. Karger AG, 2018, pp 1–10.
- 10 Islam R, Islam MRR, Tanaka T, Alam MK, Ahmed HMA, Sano H. Direct pulp capping procedures – Evidence and practice. Jpn Dent Sci Rev. 2023;59:48–61.
- Mosaddad SA, Tahmasebi E, Yazdanian A et al. Oral microbial biofilms: an update. Eur J Clin Microbiol Infect Dis. 2019;38(11):2005–2019.
- 12 Olujide S. Cohen's Pathways of the Pulp. Dent Update. 2021;48(3):248–248.
- 13 Naka S, Matsuoka D, Goto K et al. Cnm of Streptococcus mutans is important for cell surface structure and membrane permeability. Front Cell Infect Microbiol. 2022;12:994014.
- 14 Samaranayake LP. Essential microbiology for dentistry. Fifth edition. Elsevier: Edinburgh, 2018.
- 15 Larsen T, Fiehn N. Dental biofilm infections – an update. APMIS. 2017;125(4):376–384.
- 16 Lemos JA, Palmer SR, Zeng L et al. The Biology of Streptococcus mutans. Microbiol Spectr. 2019;7(1):7.1.03.
- 17 Fragkou S, Balasouli C, Tsuzukibashi O et al. Streptococcus mutans, Streptococcus sobrinus and Candida albicans in oral samples from caries-free and cariesactive children. Eur Arch Paediatr Dent. 2016;17(5):367–375.
- 18 Acevedo AM, Ray MV, Socorro M, Rojas-Sánchez F. Frequency and distribution of Mutans Streptococci in dental plaque from caries-free and caries-affected Venezuelan children. Acta Odontol Latinoam AOL. 2009;22(1):15–20.
- 19 Lima AR, Herrera DR, Francisco PA et al. Detection of Streptococcus mutans in symptomatic and asymptomatic infected root canals. *Clin Oral Investig.* 2021;25(6):3535–3542.
- 20 Thimmegowda U, Belagatta V, Chikkanarasaiah N, Bilichodmath S.

Identification and Correlation of Streptococcus mutans and Streptococcus sanguinis in Caries-active and Caries-free Children: A PCR Study. Int J Clin Pediatr Dent. 2023;16(1):9–15.

- 21 Havsed K, Stensson M, Jansson H et al. Bacterial Composition and Metabolomics of Dental Plaque From Adolescents. Front Cell Infect Microbiol. 2021;11:716493.
- 22 Oho T, Yamashita Y, Shimazaki Y, Kushiyama M, Koga T. Simple and rapid detection of Streptococcus mutans and Streptococcus sobrinus in human saliva by polymerase chain reaction. Oral Microbiol Immunol. 2000;15(4):258–262.
- 23. Adiningrat A, Fadhlurrahman AG, Rismawanto Y et al. The effects of UV light wavelength and distance differences in viral genetic material integrity. Semarang, Indonesia, 2024, p 060001.
- 24 Zhang Y, Fang J, Yang J et al. Streptococcus mutans- associated bacteria in dental plaque of severe early childhood caries. J Oral Microbiol. 2022;14(1):2046309.
- 25 Dinis M, Agnello M, Cen L et al. Oral Microbiome: Streptococcus mutans/Caries Concordant-Discordant Children. Front Microbiol. 2022;13:782825.
- 26 Fatmawati DWA. Hubungan Biofilm Streptococcus mutans Terhadap Resiko Terjadinya Karies Gigi. StomatognaticJKG Unej. 2011;8(3):127–130.
- 27 Krzyściak W, Jurczak A, Kościelniak D, Bystrowska B, Skalniak A. The virulence of Streptococcus mutans and the ability to form biofilms. Eur J Clin Microbiol Infect Dis. 2014;33(4):499–515.
- 28 Gunawan HA, Soeherwin M. Effect of Anchovy Substrate of Stolephorus baganensis on Mutans Streptococci Isolated from Human Harbouring Species. Stomatognatic JKG Unej. 2010;7(3):19–22.
- 29 Yang Y, Liu S, He Y, Chen Z, Li M. Effect of LongZhang Gargle on Biofilm Formation and Acidogenicity of Streptococcus mutans In Vitro. BioMed Res Int. 2016;2016:1–8.
- 30 Klein MI, Duarte S, Xiao J, Mitra S, Foster TH, Koo H. Structural and Molecular Basis of the Role of Starch and Sucrose in Streptococcus mutans Biofilm Development. Appl Environ Microbiol. 2009;75(3):837–841.
- 31 Aas JA, Griffen AL, Dardis SR et al. Bacteria of Dental Caries in Primary and Permanent Teeth in Children and Young Adults. J Clin Microbiol. 2008;46(4):1407– 1417.

- 32 Marsh PD. Dental plaque as a biofilm and a microbial community – implications for health and disease. BMC Oral Health. 2006;6(S1):S14.
- 33 Richert R, Ducret M, Alliot-Licht B, Bekhouche M, Gobert S, Farges J. A critical analysis of research methods and experimental models to study pulpitis. Int Endod J. 2022;55(S1):14–36.
- 34 Korona-Glowniak I, Skawinska-Bednarczyk A, Wrobel R et al. Streptococcus sobrinus as a Predominant Oral Bacteria Related to the Occurrence of Dental Caries in Polish Children at 12 Years Old. Int J Environ Res Public Health. 2022;19(22):15005.
- 35 Johansson I, Witkowska E, Kaveh B, Lif Holgerson P, Tanner ACR. The Microbiome in Populations with a Low and High Prevalence of Caries. J Dent Res. 2016;95(1):80–86.

- 36 Young DA, Nový BB, Zeller GG et al. The American Dental Association Caries Classification System for Clinical Practice. J Am Dent Assoc. 2015;146(2):79–86.
- 37 AlEraky DM, Madi M, El Tantawi M et al. Predominance of non-Streptococcus mutans bacteria in dental biofilm and its relation to caries progression. Saudi J Biol Sci. 2021;28(12):7390–7395.
- 38 Mattos-Graner RO, Klein MI, Smith DJ. Lessons Learned from Clinical Studies: Roles of Mutans Streptococci in the Pathogenesis of Dental Caries. Curr Oral Health Rep. 2014;1(1):70–78.
- 39 Pitts NB, Zero DT, Marsh PD et al. Dental caries. Nat Rev Dis Primer. 2017;3(1):17030.